



Frequently Asked Questions for Ultra BL21 (DE3) and Ultra BL21 (DE3) pLysS Competent Cells

About Your BL21 Competent Cells

1. How can we easily distinguish strain types when looking at your box?

Different strains are clearly labeled as such. While we do not label the aluminum seal affixed to the plate product, there are other distinguishing labels that make identification easy. Labeling the aluminum seal would prevent the piercing characteristic.

2. How are the E. coli strains derived?

These strains are the standard BL21 (DE3) strains initially developed by the Brookhaven National Laboratories. We have not introduced any additional modifications.

3. What are the genotypes and relative advantages of Ultra BL21 (DE3) and Ultra BL21 (DE3) pLysS Competent Cells?

Ultra BL21 (DE3): F⁻ ompT hsdS_B (r_B⁻ m_B⁻) gal dcm (DE3)

Ultra BL21 (DE3) pLysS: F⁻ ompT hsdS_B (r_B⁻ m_B⁻) gal dcm (DE3) pLysS (Cam^R)

Table with 2 columns: Genotype, Advantage. Rows include DE3, pLysS, CamR, ompT, and hsdSB (rB- mB-).

4. For what purpose(s) is your Ultra BL21 Competent Cell bacterial strain used?

These cells are ideal for high-level protein expression since they lack both ompT and lon proteases.

5. What efficiencies do your ultra competent cells attain?

Edge BioSystems guarantees >2 x 10^8 colonies / µg pUC19, although transformation efficiencies are usually higher. As with any other competent cell, the transformation efficiency will decrease with larger plasmids or ligated DNA.



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6. What are the advantages and disadvantages of using *E.coli* for protein expression?

Most recombinant proteins can be cloned and expressed in *E.coli*. The use of *E.coli* for protein expression is well documented for its advantages of low cost, easy transformation and fermentation, and high protein yields. However, solubility may be an issue, since some proteins are insoluble and aggregate in inclusion bodies. It is important to note that the following factors can affect expression levels and/or solubility: growing temperature, concentration of inducer (IPTG), host strain, protein size and structure, and toxicity.

7. Can I use my BL21 Ultra Competent Cells for non-research purposes?

The Ultra BL21 (DE3) pLysS Competent Cells are based on the T7 expression system. This is technology developed at Brookhaven National Laboratory under contract with the U.S. Department of Energy. Consequently, U.S. patents assigned to Brookhaven Science Associates (BSA) protect this technology.

These materials are to be used by noncommercial entities for research purposes only. Commercial entities require a license from BSA. You may refuse these cells by returning the enclosed materials unused.

To obtain information about licensing, please contact the Office of Intellectual Property and Partnerships, Brookhaven National Laboratory, Building 475D, Upton, NY 11973 (telephone: 631-344-7134 or fax: 631-344-3729).

8. Which BL21 strain best fits my needs?

The following chart can be used as a general guideline:

Competent Cell Strain	Description	Application
BL21 (DE3)	(DE3) indicates that the host is a lysogen of λ DE3 and therefore carries a chromosomal copy of the T7 RNA polymerase gene under control of the <i>lacUV5</i> promoter.	Recombinant protein production non-toxic to <i>E.coli</i>
BL21 (DE3) pLysS	Coupled with the response above, the addition of pLysS indicates plasmid encoding small amounts of T7 lysozyme, which is a natural inhibitor of T7 RNA polymerase and lowers the basal levels of recombinant protein before induction.	In addition to the above information, recombinant proteins can affect cell growth and viability. In the presence of detergents, small levels of lysozyme will help break the cell wall, which facilitate cell lysis once the protein is already expressed.

Protein Expression

1. Is it possible to directly clone, propagate and express into Ultra BL21 Competent Cell strains?

Yes. Edge Biosystems' Ultra BL21 Competent Cells strains have a transformation efficiency of $>2 \times 10^8$ making it possible to eliminate the intermediate step of cloning into a different strain of *E.coli*, purifying the DNA and then transforming into a BL21 strain with lower transformation efficiency.



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2. Are Ultra BL21 Competent Cells good for protein expression?

Yes, Ultra BL21 Competent Cells are identical to standard BL21. BL21 cells naturally lack *ompT* and *lon* proteases improving stability of synthesized proteins. BL21 (DE3) expresses T7 polymerase after induction with IPTG, therefore promoting the transcription of genes under the T7 promoter (for example, those in pET plasmids).

3. What proteins are the Ultra BL21 Competent Cells good at expressing?

That is difficult to predict. Protein levels as well as solubility will vary from protein to protein. In general, long proteins are much more difficult to express than shorter proteins. Also, human proteins that contain clusters of codons rarely used in *E.coli* may have a tendency to give lower yields and/or truncated products.

4. How is expression controlled?

Our Ultra BL21-derived Competent Cell strains are designed for high-level protein expression using T7 RNA polymerase-based expression systems. Our Ultra BL21 (DE3) pLysS Competent Cell strain provides tighter control for expression of toxic proteins.

5. How can we best express toxic proteins?

Proteins toxic to *E.coli* are better expressed in BL21 containing the pLysS plasmid. The pLysS plasmid will express low levels of lysozyme that will bind to T7 polymerase, therefore inhibiting transcription. This will lower the basal expression of the protein during the pre-induction growth. If the protein is extremely toxic, then it is better expressed in a BL21 (DE3) pLysE strain or using a combination of a BL21 (DE3) pLysS strain and a T7 *lac* promoter.

Ligation

1. Does ligated DNA need to be diluted before adding it to Ultra BL21 Competent Cells?

No, ligated DNA does not require dilution prior to transformation.

2. How is transformation efficiency affected by the amount of DNA used during ligation?

The more DNA used, the more efficient the ligation will be. The ratio of vector to inserted DNA can be critical for obtaining high-efficiency ligations. A molar excess of insert to vector may yield higher efficiency ligations when subcloning inserts into plasmid vectors. However, an equal or greater ratio of vector to insert may be preferred when performing library construction into plasmid vectors.

3. How does transformation efficiency of supercoiled pUC19 compare with cut/ligated DNA?

Material	Transformation Efficiency	% Efficiency Relative to Supercoiled pUC19
Supercoiled pUC19	1.8×10^7	100
Single cut pUC19	6.8×10^6	37.8
Double cut pUC19	2.4×10^6	13.3





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Note: This experiment was done using cells with low transformation efficiency. Consequently, the efficiency is usually about ten fold higher.

Induction

1. Do higher IPTG concentrations yield better expression results?

The optimal concentration of IPTG may vary from protein to protein. In other words, a concentration that works well for one protein may be too high for another, therefore resulting in insolubilization. In general, 0.4mM IPTG provides full induction of genes under the T7 promoter and 1mM is recommended for full induction of genes under the T7 *lac* promoter.

2. What promoter do the Ultra BL21 Competent Cells use?

Ultra BL21 (DE3) Competent Cells express the T7 polymerase under induction with IPTG. Plasmids containing the gene of interest under the control of a standard T7 promoter or a variant of it (for example, a T7-*lac* promoter) can be used with this system.

Freezing and Thawing

1. How should I store my competent cells when they arrive?

Competent cell stability in terms of transformation efficiency is guaranteed by Edge BioSystems for at least 3 months, when properly stored at -70°C. Upon receipt of your cell shipment, cells should immediately be placed at -70°C to ensure optimal activity.

Protein Yield Potential

1. Is it possible to predict protein yields by the strain?

No. Protein levels depend on many factors, including the protein that is expressed.

Quality Control

1. What quality control can be performed to test the transformation efficiency of the ultra competent cells?

We calculate the transformation efficiency of our ultra competent cells with supercoiled pUC19 and test the presence of the pLysS plasmid in the case of Ultra BL21 (DE3) pLysS Competent Cells by plating them in the presence of chloramphenicol.

Growth Conditions

1. How does the growth of Ultra BL21 Competent Cell strains differ from other BL21 strains currently on the market?

There is no difference from standard BL21. Some BL21 strains containing mutations will have slower growth.

Transformation and Recovery

1. How important is SOC medium when using ultra competent bacterial cells?



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An incubation of 45-60 minutes with SOC lets bacteria recover after transformation. Our transformation efficiencies are calculated using a standard 60-minute incubation with SOC after the heat shock step. If SOC recovery is omitted, the transformation efficiency may decrease up to 10-fold for ampicillin-resistant plasmids and up to 100-fold for kanamycin-resistant plasmids.

2. What formulation of SOC medium works best with this application?

Standard SOC medium can be prepared as follows:

<u>Ingredients</u>	<u>Final Concentration</u>
Yeast Extract	5g/l
Tryptone	20g/l
Sodium Chloride	10mM
Potassium Chloride	2.5mM
<u>Autoclave, then add:</u>	
Glucose	20mM
Magnesium Chloride	10mM
Magnesium Sulfate	10mM

3. Can I recover my cells in less than 1 hour (as indicated in the recommended protocol)?

When transforming with an ampicillin-based plasmid, 15-30 minutes of recovery will give about 70% of the standard efficiency. For kanamycin-based plasmids, 30 minutes of recovery will give about 50% of the standard efficiency, and 45 minutes recovery will give efficiency similar to 1 hour of recovery.

4. Can I add SOC directly to the 1.5ml tube of competent cells?

SOC added directly to the 1.5ml tube of competent cells followed by incubation at 300 rpm at 37°C will lower the transformation efficiency to about 40% of the standard efficiency.

5. Can I transform the 96-well plate using a thermocycler?

We have tested the transformation efficiency of our plates using a thermocycler with a ramp of 1.5°C per second (with actual times of 28 seconds from 0°C to 42°C and 110 seconds from 42°C to 0°C) and the transformation efficiency was 40% of the standard efficiency. Using 2 thermocyclers and transferring the plate by hand, the transformation efficiency was 60% of the standard efficiency. In both cases, the cells were recovered in 0.5ml of SOC in a 2ml deep well plate at 300 rpm, 37°C.

Transformation: Natural and Artificial

1. What does “transformation” really mean?

Transformation is the process by which naked DNA is introduced into cells. It also can happen naturally at a low frequency.

2. What exactly are “competent cells?”

These are cells that have been physically manipulated to increase their transformation efficiency.

3. What is “transformation efficiency?”

Transformation efficiency refers to the number of cells that are transformed by 1µg DNA, usually a supercoiled plasmid.





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4. How do I calculate the transformation efficiency?

$(\text{cfu on control plate}) / (\text{ng of uncut vector}) \times (10^3 \text{ ng} / \mu\text{g}) \times (\text{final dilution}) = \text{cfu} / \mu\text{g DNA}$

(Note: cfu = colony forming units)

5. How are cells made competent?

Cells can be made competent either chemically or by electroporation. Chemical competency usually involves treatment with divalent cations at low temperatures, followed by a quick cold-heat transfer during transformation. Electroporation involves the removal of salts that may cause “arching” during the electrical shock. Both methods provide cells that can be frozen for storage.

