



Troubleshooting Guide for PERFORMA[®] DTR V3 96-Well Short Plates

PROBLEM	POSSIBLE CAUSE	SUGGESTED SOLUTIONS
Signal Intensity low	Less than 10 μ l of sequencing reaction loaded	Adjust the sequencing reactions to 10 μ l before loading on the plate
	Centrifuge conditions were not optimal	<ul style="list-style-type: none"> • Recalculate rpm to achieve 850 x g for your centrifuge • Change pre-spin time to 3 min at 750 x g or decrease pre-spin time to 2-2.5 min at 850 x g • Centrifuge samples for 5 min at 850 x g • If adjustable, use fast acceleration centrifugation mode
	Sequencing reaction conditions were not optimal	Adjust sequencing reaction parameters; consult sequencing reagent manufacturer's recommendations
	Sample volume too low for sequencer	Add deionized water or sample buffer to purified reaction before loading on the sequencing instrument
Dye blobs	Too large a volume was loaded	<ul style="list-style-type: none"> • Load no more than 15 μl • If using > 3 μl BigDye[®] v 3.1 per 15 μl reaction, consider loading only 10 μl on the Performa[®] DTR V3 96-Well Short Plate • Consider using Performa[®] DTR Standard Plate for sample volumes > 15 μl, or sequencing reactions containing more than 3 μl BigDye[®]
	The concentration of BigDye [®] v. 3.1 was too high (i.e. More than 3 μ l BigDye [®] v 3.1 per 15 μ l reaction was loaded)	If the sequencing reaction conditions cannot be altered, consider loading only a portion of the reaction for cleanup and sequencing.



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Dye blobs cont...	Centrifuge conditions were not optimal	<ul style="list-style-type: none"> • Be certain that the pre-spin is for 3 min at 850 x g • Recalculate rpm to achieve 850 x g for your centrifuge conditions • Decrease sample spin to 3 minutes at 850 x g
	Samples were not loaded properly	Load the samples slowly, drop-wise to the center of the gel matrix
	Signal intensity too high	Apply only a portion of sequencing reaction to the plate; high signal strengths exaggerate the effect of small quantities of dye terminator in the purified reaction
	Sequencing reaction conditions were not optimal	Adjust sequencing reaction parameters; consult sequencing reagent manufacturer's recommendations
Truncation of 5' sequence	Centrifuge conditions were not optimal	<ul style="list-style-type: none"> • Recalculate rpm to achieve 850 x g for your centrifuge • Change pre-spin time to 3 min at 750 x g or decrease pre-spin time to 2-2.5 min at 850 x g • Centrifuge samples for 5 min at 850 x g
	Dye blobs present	See Dye blobs above
	Less than 10 μ l of sequencing reaction loaded	Adjust the sequencing reaction to 10 μ l before loading on the plate
	Sequencing reaction conditions not optimal	Adjust sequencing reaction parameters; consult sequencing reagent manufacturer's recommendations



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Recovery volume higher than expected	Centrifuge conditions were not optimal	<ul style="list-style-type: none"> • Be certain that the pre-spin is performed for 3 min at 850 x g • Recalculate rpm for your centrifuge conditions
	More than 15 μ l of sequencing reaction loaded	Decrease volume of sequencing reaction loaded to 10-15 μ l
	Plate was stored incorrectly. If you observe a high recovery volume after the first spin, it is likely the plate has been frozen	<ul style="list-style-type: none"> • Store sealed plates at 4°C • Do not allow plates to freeze
Recovery volume lower than expected	Centrifuge conditions were not optimal	<ul style="list-style-type: none"> • Recalculate rpm for your centrifuge conditions • Spin samples for at least 5 min at 850 x g
	Less than 10 μ l of sequencing reaction loaded	Increase volume of sequencing reaction to 10 -15 μ l
	Plates have not been stored correctly. If you observe a low recovery volume after the first spin, it is likely the plate has been dehydrated	<ul style="list-style-type: none"> • Store plates at 4°C • Plates sealed in original bag can be left at room temperature for several days but should be stored at 4°C

For additional troubleshooting assistance, please contact Edge BioSystems at:

Telephone: (800) 326-2685 or (301) 990-2685	Fax: (301) 990-0881
Email: info@edgebio.com	Web: http://www.edgebio.com